Registration form of Stranded Cetaceans in the WCR

Stranded cetacean data

Location:_____

_____

_____

Lat: _____˚N Long _____º W.

Date of the stranding: _____

Death date: _____

Data collection date: _____

Number of individuals: _____

Likely death cause: _____

Condition of the individual: (1) alive; (2) recent death; (3) decomposing (organs intact, little odor, and little swelling); (4) advanced decomposition (unrecognizable organs, body intact); (5) very advanced decomposition (dry body or skeleton only

_____

_____

Missing parts: _____

(Write down the condition of the specimen in its location and in its subsequent rescue).

Description: color, location of scars, wounds, parasites, and other interesting details.

Head:_____

_____

Dorsal Region: _____

_____

Sides:_____

_____

Abdominal Region: _____

_____

Dorsal fin: _____

Pectoral fins: (dorsal)_____

(ventral):_____

Caudal fin: (dorsal)_____

(ventral) _____

Very important: Use the drawings on this form to detail the characteristics of the specimen, paying attention to its colour, head shape, pectoral, dorsal and caudal fins. On the inside of this form you can make sketches or outlines of the head and fins. Indicate missing parts of the specimen with dotted lines on your drawing

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_____ _____

Register number Sex

_____ _____

Total length Weight

Species: _____

Teeth: UL_____ UR_____

LL_____ LR_____

Baleens : UL_____ UR_____

Reported by: _____

Address:_____

_____Phone:_____

Data taken by: _____

_____

_____Phone: _____

Weather and tidal conditions at the time of the stranding:_____

_____

Body Orientation: _____

If it is alive, indicate the presence or not of predators or enemies, and behavior of the specimen before and after stranding:_____

_____

_____

Alive Animal		**Outcome**	
Healthy	_____	Refloated and survive	_____
Emaciated	_____	Refloated and die	_____
Injured	_____	Intervened with meds	_____
Left alone	_____	Kept public away	_____
Sick in water	_____	Swam away	_____
Injured in water		Died	

Samples Obtained: at least they must be obtained from:

Mysticetes (baleen whales): two central baleen plates or ear bone _____

Odontocete (with teeth): 4-5 consecutive teeth from the central part , whole head, skull or fins

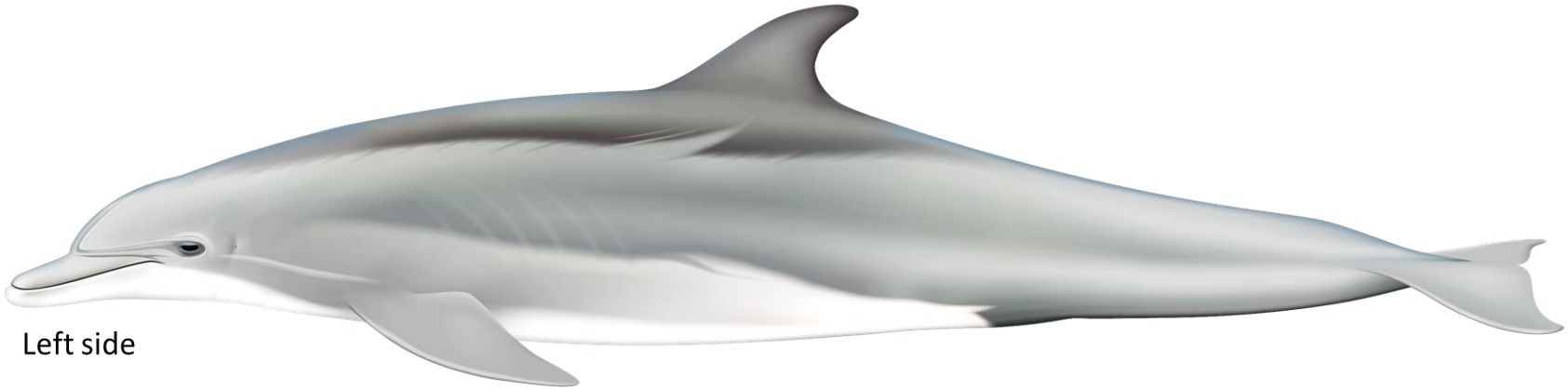
Use of the individual: _____

Destination of the samples: _____

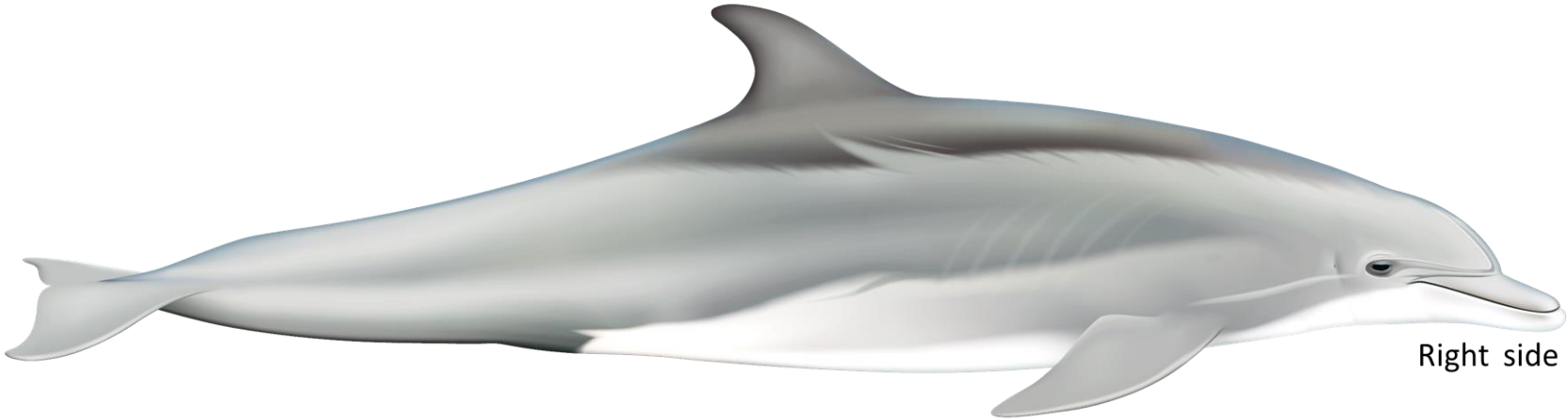
_____

Use the inner part of the form to draw the outline (silhouette) of the pectoral (indicating right or left of the animal), dorsal and caudal fins. Place on paper under the fins and trace the outline with a pencil in a vertical position. If possible, do the same with the head. If the animal is too big, sketch the shape of the fins and draw the distribution of colors. Don't forget to draw both pectoral fins

Draw any interesting detail, missing part, scar, parasite, net line



Left side



Right side

Comments:

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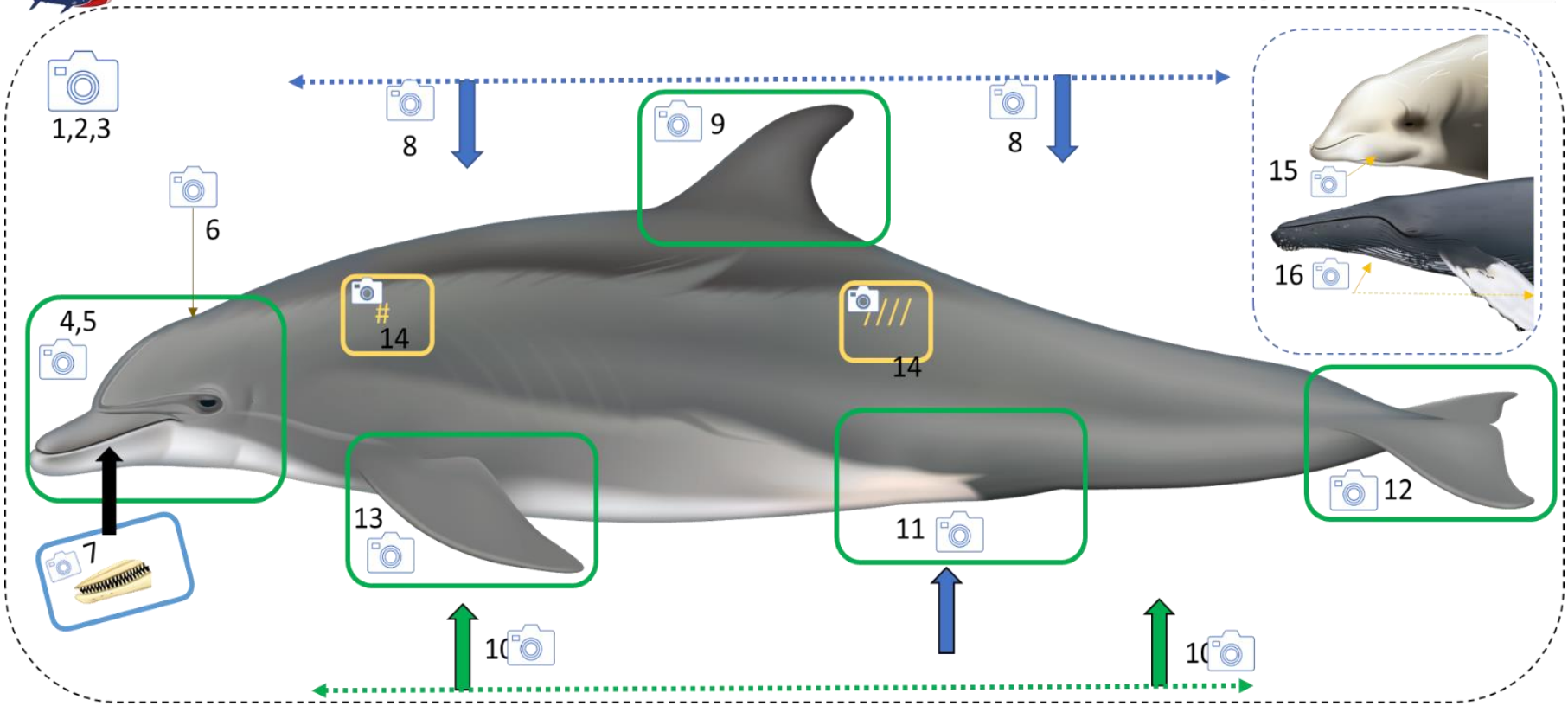
Photos to collect from a stranded dolphin



Fotos to collect. 1.Initial photo (complete body before moving the animal and the background). **2. Left side** (complete animal) **3. Right side** (complete animal) **4. Left head 5. Right head 6. Blowhole 7. Teeth or baleen 8. Dorsum** (back of the complete animal if posible**) 9. Dorsal fin** (look for any detail) **10. Ventrum** (belly of the complete animal if posible**) 11. Genital área 12. Fluke 13.Pectoral flipper** (take both together and then each separate). **14. Wounds or marks. Special measurments: 15. Beaked whales: throat grooves 16. Whales: Throat grooves** (try to photograph the number and the extension).

Comments:

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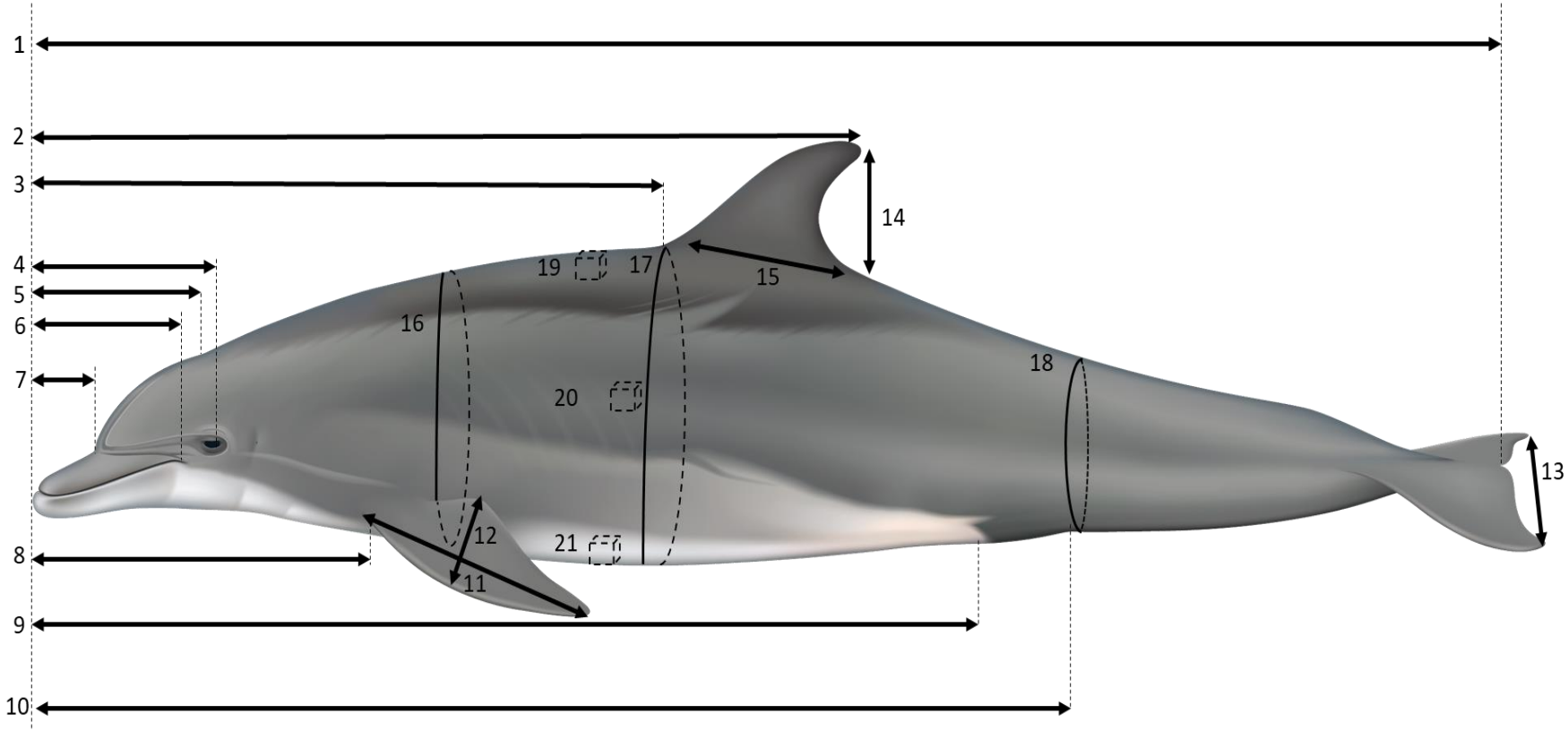
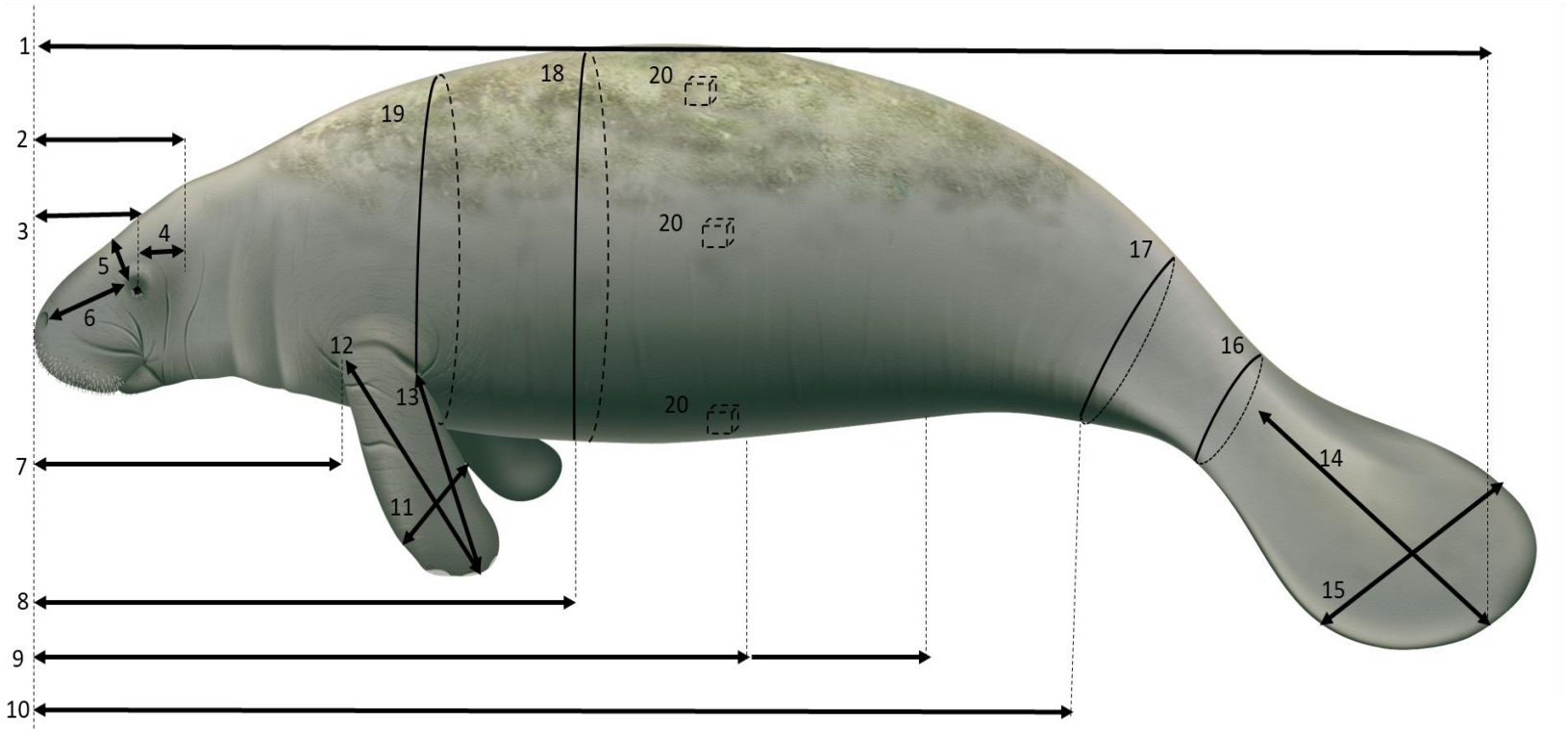
MEASURING STRANDED CETACEANS



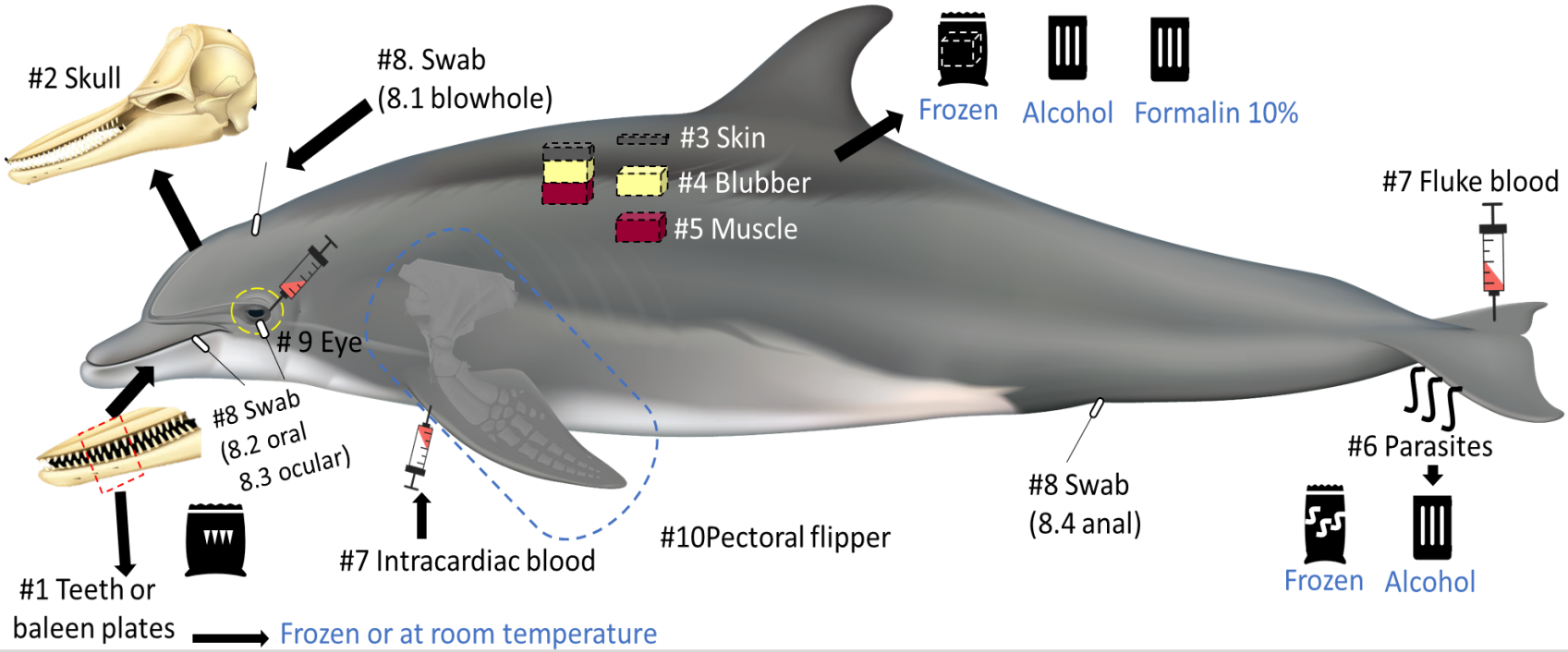
Basic measurments in stranded cetaceans. 1. Snout to fluke notch. **2.** Snout to tip of dorsal fin. **3.**Snout to anterior insertion of dorsal fin **4.** Snout to center of the eye **5.** Snout to the blowhole **6.** Snout to the angle of the mouth **7.** Snout to melon **8.** Snout to the anterior insertion of flipper. **9.**Snout to center of genital apertura **10.** Snout to the center of the anus **11.** Flipper length **12.** Flipper width **13.** Fluke width **14.** Dorsal fin height **15.** Dorsal fin base **16.**Girth: axillary **17.** Girth: maimun (specify location) **18.** Girth: at anus **19.** Blubber thickness: dorsal (anterior and lateral to dorsal fin) **20.** Blubber thickness : lateral at mid-length (anterior and lateral to dorsal fin) **21.** Blubber thickness: ventral at mid-lenght.(anterior and lateral to dorsal fin) (Adapted from Geraci & Lounsbuty,2005).

MEASURING STRANDED MANATEES



Basic measurments in stranded manatees. 1. Tip of snout to tip of fluke. **2** Tip of snout to external ear. **3.** Tip of snout to center of the eye **4.** Center of the eye to ear **5.** Distance between centers of eyes **6.** Center of eye to center of nostril (same side) **7.** Tip of snout to the anterior insertion of flipper. **8.** Tip of snout to center of the umbilicus **9.** Tip of snout to center of genital aperture **10.** Tip of snout to the center of the anus **11.** Maximun width of flipper **12.** Flipper length, anterior insertion to tip width **13.** Flipper length, axilla to tip **12.** Fluke width **13.** Dorsal fin height **14.** Base of fluke to posterior tip **15.** Maximun width of fuke. **16.**Girth at fluke base **17.** Girth at anus**. 18** Girth at umbilicus **19.** Girth at axilla **20.** Thickness of skin: dorsal (anterior, lateral, ventral). (Adapted from Geraci & Lounsbuty,2005[XX]).

Basic samples to collect and preserve (DDC 2-3)



Basic samples to collect from a stranded cetacean 1. Teeth sample: Count the total teeth (or baleen) and empty sockets and collect 4-5 teeth or baleen (from middle part of the jaw), store frozen or dry in a bag. **2. Skull** can be preserved by different methods (if collected try to get a brain and cerebro espinal fluid samples). **Samples #3 skin, #4 blubber, # 5 muscle** cut a square of 5x5 cm using a knife or blade and tweezers and separate them. The samples for formalin should have maximun 1 cm thick. The alcohol could be 70% ideally at 95% (even vodka and gin works!). They can be frozen directly. Blubber ideally should be wrap first with aluminium paper foil and then put it in a bag.**6.External parasites.** Storage in alcohol at room temperature and/or frozen. **7. Complete blood** can be collected ideally intracardiac untill 16 hours after death. For serum deposit it in red cap blood tubes. **8.Swabs:** Sterile swabs from the blowhole, oral, ocular and anal can be collected for disease diagnoses .**9. Eye:** both eyes can be collected for serology, and histopathology **10.Pectoral flipplers**: can be collected and frozen for later interpretation of age class by Xrays. **IMPORTANT: remember to label each bag with the case identification and date!**

Comments:

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Organ to sample **(cause of death cannot be determined!)*	**Analyses:** ✓ life history **(Max DDC=5)** ✓ DNA **(Max DDC=3)** ✓ Biomarkers **(Max DDC=1)** ✓ microbiology (virus, bacteria, biotoxins) **(Max DDC=2-3)** ✓ stable isotope &fatty acids **(Max DDC=5)** ✓ serology **(Max DDC=2)** **Storage:** plastic bag in Freezer (-20 to -80°C)				**Analyses**: contaminants **(Max DDC=3)** **Storage:** wrapped in aluminum foil and put in plastic bags in freezer (-20 to -80°C)		**Analyses**: histology (pathology & reproduction) **(Max DDC=3)** **(no more than 1 cm thick!)** **Storage:** plastic container with 10% buffered formalin (9 times liquid versus tissue), at room temperature		**Analysis:** parasitology **(Max DDC=4)** **Storage:** Plastic vial with alcohol 10% in freezer (-20 to -80°C), and room temperature	
	What for	Container	Sample Size	Number of samples	Sample Size	Number of samples	Sample Size	Number of samples	Sample size	Number of samples
Teeth	Life history	Sterile plastic bag	5 teeth collected in the middle of the jaw							
Skull	Life history	Plastic bag	All							
Skin	DNA Microbiology	Sterile plastic bag	5x5 cm	2 ☐			3x3 x **0.5 cm**	1 ☐		
Blubber	Stable isotopes & fatty acids Microbiology	Sterile plastic bag	5x5x5 cm	2 ☐	5x5x5 cm	1 ☐	5x5x**1 cm**	1 ☐	Several parasites	2 ☐
Muscle	Stable isotopes & fatty acids Microbiology	Sterile plastic bag	5x5x5 cm	2 ☐	5x5x5 cm	1 ☐	5x5x**1 cm**	1 ☐	Several parasites	2 ☐
External parasites (mites & lice)									Several parasites	2 ☐
Complete blood (if less than 16 hours of death)	Microbiology Serology	Blood collector tube (red cap)	5-10 ml	2 ☐						
Eye-fluid (alternative to blood if carcass not fresh)	Serology	Sterile vial 1.5 ml	3 ml	1 ☐						
Eye and ocular swab	Microbiology Histology	Sterile plastic bag	All organ (1 eye)	1 ☐ 1 from eye ☐			(2^{nd} eye) (inject formalin)	1 ☐		
Respiratory swab	Microbiology	Sterile swab stick	1 from blowhole ☐							
Digestive swabs	Microbiology	Sterile swab stick	1 from anus ☐							
Scapula	Life history		All organ	2 ☐						

List of samples to collect for an extensive data collection

Organ to sample	**Analyses:** ✔ life history **(Max DDC=5)** ✔ DNA **(Max DDC=3)** ✔ Biomarkers **(Max DDC=1)** ✔ microbiology (virus, bacteria, biotoxins) **(Max DDC=2-3)** ✔ stable isotope &fatty acids **(Max DDC=5)** ✔ serology **(Max DDC=2)** **Storage:** plastic bag in Freezer (-20 to -80°C)				**Analyses**: contaminants **(Max DDC=3)** **Storage:** wrapped in aluminum foil and put in plastic bags in freezer (-20 to -80°C)		**Analyses**: histology (pathology & reproduction) **(Max DDC=3) (no more than 1 cm thick!)** **Storage:** plastic container with 10% buffered formalin (9 times liquid versus tissue), at room temperature		**Analysis:** parasitology **(Max DDC=4)** **Storage:** Plastic vial with alcohol 10% in freezer (-20 to -80°C), and room temperature	
	What for	Container	Sample Size	Number of samples	Sample Size	Number of samples	Sample Size	Number of samples	Sample Size	Number of samples
Tonsils	Microbiology	Sterile plastic bag	5x5 cm	1 ☐			3x3x1cm	1 ☐		
Lung		Sterile plastic bag	5x5 cm	1 in upper location ☐ 1 in lower location ☐			3x3x1cm	1 ☐	Several parasites	2 ☐
Lymph nodes		Sterile plastic bag	Half	1 ☐			half	1 ☐		
Heart/ cardiac muscle		Sterile plastic bag	5x5 cm	1 ☐			3x3x1cm	2 in different regions ☐		
Liver		Sterile plastic bag	5x5 cm	2 ☐	5x5 cm	1 ☐	3x3x1cm	1 ☐	Several parasites	2 ☐
Pancreas		Sterile plastic bag	5x5 cm	1 ☐			3x3x1cm	1 ☐		
Kidney		Sterile plastic bag	5x5 cm	1 ☐	5x5 cm	1 ☐	3x3x1cm	1 ☐		
Adrenal glands		Sterile plastic bag	5x5 cm	1 ☐	5x5 cm	1 ☐	3x3x1cm	1 ☐		
Spleen		Sterile plastic bag	half	1 ☐			half	1 ☐	Several parasites	2 ☐
Thyroid gland		Sterile plastic bag	5x5 cm	1 ☐	5x5 cm	1 ☐	3x3x1cm	1 ☐		
1r. Stomach		Sterile plastic bag	5x5 cm	1 ☐			3x3x1cm	1 ☐		
2d. Stomach		Sterile plastic bag	5x5 cm	1 ☐			3x3x1cm	1 ☐		

Organ to sample	**Analyses:** ✔ life history **(Max DDC=5**) ✔ DNA **(Max DDC=3)** ✔ Biomarkers **(Max DDC=1)** ✔ Microbiology (virus, bacterias, biotoxins) **(Max DDC=2-3)** ✔ Stable isotope &fatty acids (**Max DDC=5)** ✔ Serology **(Max DDC=2)** **Storage:** plastic bag in Freezer (-20 to -80°C)				**Analyses**: contaminants **(Max DDC=3)** **Storage:** wrapped in aluminum foil and put in plastic bags in freezer (-20 to -80°C)		**Analyses**: histology (pathology & reproduction) **(Max DDC=3)** **Storage:** plastic container with 10% buffered formalin (9 times liquid versus tissue), at room temperature		**Analysis:** parasitology **(Max DDC=4)** **Storage:** Plastic vial with alcohol 10% in freezer (-20 to -80°C), and room temperature	
	What for	Container	Sample Size	Number of samples	Sample Size	Number of samples	Sample Size	Number of samples	Sample Size	Number of samples
Stomach content	Microbiology	Sterile vial	10 ml	1 ☐					Several parasites	1 ☐
	Diet from prey analysis	Sterile plastic bag	Entire content ☐							
Intestine	Microbiology	Sterile plastic bag	5x5 cm	2-3 regions ☐			5x5x1 cm	2-3 regions ☐	Several parasites	1 ☐
Faeces		Sterile vial	1-10 ml	1 ☐					Several parasites	1 ☐
Testicles/Ovary		Sterile plastic bag	half	1 ☐			half	1 ☐		
Milk		Sterile plastic vial	1-10 ml	1 ☐						
Placenta		Sterile plastic bag	5x5 cm	1 ☐	5x5 cm	1 ☐	5x5x1 cm	1 ☐		
Fetal tissue (liver,kidney,lung, brain, stomach content)		Sterile plastic bag	3x3 cm	1 ☐	3x3 cm	1 ☐	5x5x1 cm	1 ☐		
Mammary gland		Sterile plastic bag	5x5 cm	1 ☐			5x5x1 cm	1 ☐		
Brain **Cerebro spinal fluid**		Sterile plastic bag Syringe or 1.5 vial	5x5x5 cm 0.1- 5 ml	2-3 regions ☐ 1-3 ☐			Half brain with cuts in the tissue so the formalin can penetrate adequately	1 ☐		

Notes: